

Short Communication

J Indian bot Soc Vol 72 (1993) 183-184

INHIBITION OF AFLATOXIN PRODUCTION ON MAIZE AND MUSTARD CROP VARIETIES BY AQUEOUS EXTRACT OF *EUPHORBIA HIRTA* L.

G. PRASAD AND S. KUMAR

University Department of Botany Bhagalpur University, Bhagalpur-812 007

Key Words: Aflatoxin, mustard, maize.

The antiaflatoxicity of aqueous extract of *Euphorbia hirta* L. was tested in the varieties of maize and mustard crops. The maximum inhibition of aflatoxin was observed in DC (33.42%) followed by SC (27.56%) and M₉ (19.25%). In mustard crop, 73.84% aflatoxin inhibition was recorded in Varuna and 50.56% in BR-40.

The therapeutic values of *E. hirta* is well known in curing branchial eye, stomach and kidney troubles. In addition, Santhal tribals, Bihar, largely use its juice in lactating mothers when they are suffering from milk deficiency or in the condition of failure of lactation. Antiaflatoxic property in aqueous extract of *E. hirta* is earlier described under laboratory conditions by Singh and Sinha (1985). However, Its role in crop protection against the growth of aflatoxic strains of *Aspergillus flavus* is not known so far. In the present investigation aqueous extract of *E. hirta* was tried to record its influence against aflatoxin contamination in maize and mustard crops.

Maize varieties Diara Composite (DC), Suwan Composite (SC) and M₉ and mustard varieties Varuna and BR-40 were cultivated separately in strip-strip potting with adequate irrigation and fertilizer facilities. Maize crop was cultivated during Kharif (June-Sept.) and mustard crop during Rabi (Oct.-Feb.) seasons of 1989-1990. The inoculum of toxigenic strain of *A. flavus* (SRRC-1007) was injected after seed setting in the maize cobs and by spraying spore suspension (100 spores/ml) on mustard pods. After 7 days of inoculation, aqueous extract (2:10 w/v) of *E. hirta* was sprayed on maize cobs and mustard pods. Control set was maintained with sterile distilled water. After harvest, seeds of maize and mustard were collected separately and extraction of aflatoxins was carried out by methanol-chloroform (Thomas *et al.*, 1975). Type of aflatoxins were confirmed by thin

layer chromatoplate as described by Reedy *et al.* (1970) and their quantitation by fluorodensitometer (CAMAG-TLC Scanner). The chemical confirmation of aflatoxins was done by trifluoroacetic acid and 50% sulphuric acid aqueous solution as suggested by Stack and Pohland (1975).

Aflatoxin contamination was significantly inhibited in maize and mustard crops by the treatment of *E. hirta*. The control (untreated) maize plants exhibited 2140 µg/kg aflatoxin B₁ level in variety M₉ which followed by SC (1640 µg/kg) and DC (958 µg/kg). However, treatment with *E. hirta* prevented significantly to aflatoxin elaboration which was recorded 33.42% in variety DC, 27.50% in SC and 19.25% in M₉. Untreated (control) mustard crop also exhibited high level of aflatoxin B₁ contamination after infestation of *A. flavus* toxigenic strain. The aflatoxin B₁ amount detected in variety Varuna was 1101 µg/kg and in BR-40 981 µg/kg. After treatment with *E. hirta* to mustard crops, 73.84% aflatoxin production was inhibited in variety Varuna and 50.56% in BR-40.

Maize and mustard crops exhibited high degree of aflatoxin inhibition by the treatment of *E. hirta* aqueous extract. Such inhibition possibly due to interference of active ingredients of the extract which causing interruption of growth and toxin producibility of the *A. flavus*. Earlier Singh (1981) and Singh and Sinha (1985) also suggested inhibition of aflatoxin production *in vitro* with the extract of *E. hirta*. Results also indicated the variation of toxin producibility of *A. flavus* (SRRC-1007) as well as variation of toxin inhibition potentiality of *E. hirta* with regards to crop varieties. This may be attributed to the nutritional factors of the varieties. There are distinct variation in the nutritional composition of two varieties. Thompson *et al.* (1984) and Kumar (1990) have also found such type of variation of

aflatoxin production between two varieties. Our findings thus, may suggest that, if inhibitory factor of *E. hirta* could be examined that can be advantageous for preventing/minimizing aflatoxin contamination in maize and mustard crops.

Authors are grateful to prof. K.S. Bilgrami, Head, University Department of Botany, Bhagalpur University for providing laboratory facilities. Financial assistance received from ICAR out of PL-480 fund is also thankfully acknowledged.

REFERENCES

- Kumar Shivendra 1990 Effect of Agro-climatic conditions on Aflatoxin elaboration in mustard crops, *J Indian bot Soc* **69** 319-320.
- Reddy T V L Viswanathan & T A Venkitasubramanian 1970 Thin layer chromatography of aflatoxin, *Anal Biochem* **38** 568-571.
- Stack M E & A E Pohland 1975 Collaborative study of a method for chemical confirmation of identify of aflatoxin, *J Assoc off Anal Chem* **58** 110-113.
- Singh, Premlata 1981 *Studies on the effect of some plant extracts on aflatoxin production*. Ph D Thesis, Bhagalpur University, Bhagalpur.
- Singh P L & K K Sinha 1985 Inhibition of aflatoxin production on some agricultural commodities through aqueous plant extracts, *J Indian bot Soc* **65** 30-32.
- Sinha K K 1985 Screening of chlorophyllous plants against aflatoxin production and aflatoxin producing fungi, *J Food Sci Technol* **22** (3) 225-228.
- Thomas F, R M Eppley, & M W Trucksess & 1975 Rapid screening method for aflatoxins and zearalenone, *J Assoc Off Anal Chem* **58** 114-116.
- Thompson D L, J O Rawlings M S Zuber G A Payne & E B Lillehoj 1984 Aflatoxin accumulation in developing kernels of eight maize single crosses after inoculation with *Aspergillus flavus*, *plant Disease* **68** 465-469.